Wound healing is promoted by *Musa paradisiaca* (banana) extract in diabetic rats

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Abstract

Introduction: Impairments in wound healing commonly occur among patients with diabetes. Herbal medicines have a long history of usage in wound care management. Super green (SG) is a newly discovered natural product obtained from *Musa paradisiaca*. This study aimed to investigate the efficacy of the topical application of SG in healing surgical wounds in diabetic rats.

Material and methods: Wistar rats received a one-time intraperitoneal injection of streptozotocin to induce type 1 diabetes. Full-thickness excisional skin wounds were created on the backs of the rats. The relevant groups were topically treated with the indicated concentrations of SG or vehicle dressing throughout the study duration. Histological analysis was performed and the mRNA levels of proinflammatory cytokines were measured to evaluate the improvement of wound closure.

Results: The wound area ratio of the SG (1/6000 dilution)-treated group was greatly reduced compared to that of the vehicle-treated group. The histological analysis showed fewer inflammatory cells, accelerated re-epithelialization, and increased collagen deposition in SG 1/6000-treated wounds. The gene expression levels of tumor necrosis factor-α, interleukin-1β, and interleukin-6 were decreased and the levels of type I and type III collagen were increased after SG treatment.

Conclusions: These results show that the most therapeutically efficacious concentration of SG (1/6000 dilution) can enhance wound repair in diabetic rats. SG has the potential to be a new treatment strategy for diabetic wounds.

Key words: wound healing, diabetes, *Musa paradisiaca* extract, inflammatory cytokines.

Introduction

Diabetes mellitus (DM) is a metabolic disorder characterized by chronic hyperglycemia that currently affects over 472 million people worldwide [1]. Diabetes is often accompanied by a range of disorders, including obesity [2], atherosclerosis [3], peripheral arterial disease [4], and obstructive sleep apnea [5]. Impaired wound healing is a common complication in patients with diabetes that severely affects life quality.
Normal wound healing is a dynamic and complex process involving inflammation, re-epithelialization, neovascularization, and tissue remodeling [7]. Persistent inflammation, poor circulation and limited nutrients [8] are associated with impaired diabetic wound healing [9]. Hyperglycemia increased oxidative stress and led to development of chronic low-grade inflammation [10]. A cluster of pro-inflammatory cytokines, such as tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), and interleukin-6 (IL-6), may significantly influence the progression of inflammation in diabetes [11]. Treatments that enhance diabetic wound healing are often associated with reducing inflammatory cytokines in the diabetic wound environment [12]. Although the treatment of diabetic wounds has been greatly improved, the management of severe or chronic wounds is still challenging. Therefore, it is important to develop novel therapies for patients with wound healing difficulties.

Natural products and their phytochemicals have been used for wound healing for a long time. Natural products with wound healing properties have been reported to have anti-inflammatory, antioxidant, antibacterial and procollagen synthesis actions [13]. *Musa paradisiaca* (Linn.), commonly known as banana, belongs to the family Musaceae, which mainly grows in tropical and subtropical regions. Banana fruits and peels are rich in dietary fibers, vitamins, potassium, magnesium, and serotonin [14]. Different parts of this plant have been traditionally used for the treatment of various diseases. The fruits are used for the treatment of diarrhea, diabetes, and intestinal lesions; the leaves are used for the treatment of eczema and burns; the flowers are used for the treatment of dysentery and menorrhagia; the roots are used for anthelmintic treatment; and so on [15]. Recently, pharmacological investigations have shown that some of the bioactive phytochemicals of *Musa paradisiaca* have antioxidative stress [16], antimicrobial [17], antidiabetic [18], and wound healing properties [19]. It was recently discovered that super green (SG), a water-soluble substance extracted from the leaves, peels and stems of *Musa paradisiaca* (Linn.), seems to have antibacterial effects. The present study investigated the role of SG in the healing of surgical wounds in diabetic rats. The results will be useful for its development as a drug for the enhancement of wound healing.

**Material and methods**

**Experimental animals**

Eight-week-old male Wistar rats (250–280 g) were obtained from the National Institute of Legal National Laboratory Experimental Animal Center. The rats were maintained at a controlled temperature of 22 ±2°C on a 12-h light/dark cycle. The experimental protocols were approved by the Institutional Animal Ethics Committee of Tajen University (No. 102-22). All experiments conformed to the Guide for the Care and Use of Laboratory Animals as well as the guidelines of the Animal Welfare Act.

**Streptozotocin (STZ)-induced diabetes**

Diabetes was induced in the rats by using streptozotocin (Sigma-Aldrich, St. Louis, MO, USA), a compound that has a preferential toxicity toward pancreatic β cells. Diabetic conditions were induced by a single intraperitoneal injection of STZ at a concentration of 60 mg/kg body weight in overnight-fasted rats [20]. After three days of induction, blood samples were collected from the tail vein and measured using an automatic analyzer. Animals with fasting blood glucose levels greater than 250 mg/dl were considered diabetic.

**Dosing protocol of SG and standard drug**

SG was obtained by patented extraction technology and was provided by Natural Well Technical Ltd. Company, Taiwan. The SG stock solution was diluted 1 : 6000 (SG/6000), 1 : 3000 (SG/3000), 1 : 1000 (SG/1000) and 1 : 500 (SG/500) and mixed with ointment base to obtain a 1% SG ointment. An antimicrobial drug, chlorhexidine gluconate solution (Panion & BF Biotech Inc, Taipei, Taiwan), was mixed with the ointment base to obtain a 4% chlorhexidine gluconate (CG) ointment. Additionally, a commercial 1% Sulfasil ointment (silver sulfadiazine, Sinphar, Yilan, Taiwan) was used.

**Excision wound model and experimental design**

One week after diabetes induction, the diabetic rats were randomly classified into seven groups (n = 6): group 1 (the vehicle-treated diabetic group), group 2 (the CG-treated diabetic group), group 3 (the Sulfasil-treated diabetic group), group 4 (the SG/6000-treated diabetic group), group 5 (the SG/3000-treated diabetic group), group 6 (SG/1000-treated diabetic group), and group 7 (the SG/500-treated diabetic group). The rats were anesthetized with 2% isoflurane to minimize suffering. After the hair on the back of each rat had been removed, the skin was sterilized with 70% alcohol, and a full-thickness excisional wound (1.5 cm × 1.5 cm) was created on the dorsal skin. The drugs were applied topically once a day for 15 consecutive days of treatment. Each rat was individually caged. All rats had free access to food and water.
Wound healing analysis

Rat wounds were photographed under isoflurane anesthesia on day 0, day 3, day 7, day 10 and day 15 after induction. A standard ruler was placed on the side of the wounds. The wound areas were measured using ImageJ Software, and the area observed on day 0 was considered 100%.

Histopathological examination

At the end of the experiment, rats were anesthetized with 2% isoflurane, and the wound area was sampled for histological examination. Skin tissue samples were formalin-fixed and embedded in paraffin. The section thickness of skin tissue was 5 μm. After hematoxylin and eosin (HE) staining, wound healing was evaluated in terms of the extent of dermal inflammation, angiogenesis, presence of granulation tissue, and re-epithelialization [21]. The parameters used to assess healing were as follows: for dermal inflammation, “−” – no dermal inflammation, “+” – mild dermal inflammation, “++” – moderate dermal inflammation; and “+++” – marked dermal inflammation; for angiogenesis, “−” – no angiogenesis, “+” – mild angiogenesis (< 5 vessels per high-power field), “++” – moderate angiogenesis (6–10 vessels per high-power field), and “+++” – marked angiogenesis (> 10 vessels per high-power field); for granulation tissue thickness, “−” – none, “+” – scant, “++” – moderate, and “+++” – marked; for re-epithelialization, “−” – no re-epithelialization, “+” – < 50% of the wound, “++” – > 50% of the wound, and “+++” – complete re-epithelialization [22, 23].

Collagen deposition in diabetic wounds was assessed by Masson’s trichrome staining [24]. Collagen fibers were stained blue, and muscle and connective tissues were stained red. HE staining or Masson’s staining was performed according to the manufacturer’s protocol. Samples were observed using optical microscopy. The collagen deposition volume and other parameters were quantified using ImageJ Software. Five different fields of vision were randomly selected.

Real-time quantitative PCR detection

The expression of collagen I, collagen III, IL-1β, IL-6 and TNF-α in wound tissues in STZ-treated rats was measured using RT-PCR. Total RNA was extracted from tissue samples using the TRIzol method (Thermo Fisher Scientific, Waltham, MA, USA) and processed according to the manufacturer’s instructions. Five micrograms of RNA were reverse transcribed to cDNA using the Transcriptor First Strand cDNA Synthesis Kit (Roche, Basel, Switzerland). Amplification and detection were performed with a LightCycler 480 system (Roche). Gene expression was quantified using the relative standard curve method. β-Actin was used to normalize the RT-PCR results. The primer sequences used were as follows:

- **collagen I (α1) F:** 5'-CTGGCAAGACGGAGATGAT-3';
- **collagen I (α1) R:** 5'-AATCCAGGACACCCTGTA-3';
- **collagen III (α1) F:** 5'-CAGGAAACCCAGGAAAGG-3';
- **collagen III (α1) R:** 5'-CCAGCTACTCTATAGTCCTGG-3';
- **TNF-α F:** 5'-CACGCTTCTCTTTTCTTTT-3';
- **TNF-α R:** 5'-GCCAGAGGCGCTGATTAGAGA-3';
- **IL-6 F:** 5'-GATGAGATACACACCCGATG-3';
- **IL-6 R:** 5'-CTGCAAGCAGAACTGGTCTGT-3';
- **IL-1β F:** 5'-CAGGCCATCTTGATTGCTCA-3';
- **IL-1β R:** 5'-AGTCATCTCATTGCCACTG-3';
- **β-actin F:** 5'-CTAAGGCAACCCGTGAAAG-3';
- **β-actin R:** 5'-GCCGTGATGATGCTACACA-3'.

Statistical analysis

The data are expressed as the means ± SEM (n = 6). Statistical analysis was performed by one-way ANOVA followed by the Bonferroni post hoc test. The statistical analysis software used was SPSS 21. P < 0.05 was considered statistically significant.

Results

Effect of SG on wound healing in diabetic rats

To evaluate the effect of SG ointment on lesion area, we compared wound healing in diabetic rats treated with different drugs. In morphological microanalysis, the wounds appeared clean without signs of irritation or bleeding throughout the experiment (Figure 1A). On day 10, the topical application of SG at various dilutions and Sulfasil significantly decreased wound size compared to that in the vehicle-treated group. On day 15, the wound closure effect of SG at a dilution of 1/6000 was more effective than that of other dilutions of SG and was comparable with the effect of Sulfasil (Figure 1B). Figure 1C shows the wound area ratio of each group during the wound healing process. Healing was promoted after SG and Sulfasil treatment, and the wound area ratio of the SG/6000-treated group was more decreased between day 7 and day 10 than that of the other groups. However, CG failed to affect wound contraction compared with that in other groups.

Histopathological examination

Images of HE-stained wound sections of skin on day 15 are shown in Figure 2. Inflammatory cell infiltration (inflammation), epidermal cell regeneration (re-epithelialization), granulomas (granulation), and the number of blood vessels (angiogenesis) are presented in Table I. Inflammatory cell infiltration was observed in all diabetic groups; however, SG/6000, SG/3000 and Sulfasil treatment decreased the inflammatory effects.
Among these treatments, SG/6000 was the most effective. The number of new blood vessels in the lesions was increased in the SG/6000–, SG/3000– and Sulfasil-treated groups. To evaluate the kinetics of wound closure, the number of granulomas was measured. The SG/6000– and Sulfasil-treated groups exhibited more granulomas than the other groups. Moreover, the SG/6000-treated group showed higher re-epithelialization levels than those of the other groups. After treatment with CG, the wound size of the STZ-treated diabetic rats was not obviously improved.

Analysis of collagen deposition

Total collagen content was analyzed using Masson’s trichrome staining. Our results indicat-
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The parameters used to assess the healing of excision wounds in the diabetic rats were evaluated as follows: dermal inflammation (inflammation), neovascularization (angiogenesis), epidermal epithelial cell regeneration and healing (re-epithelialization) and granulation tissue (granulation).

Table 1. Histological comparison of excision wound values per treatment

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Inflammation</th>
<th>Angiogenesis</th>
<th>Granulation</th>
<th>Re-epithelialization</th>
</tr>
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<tbody>
<tr>
<td>Vehicle</td>
<td>+++</td>
<td>+</td>
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<tr>
<td>CG</td>
<td>+++</td>
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<tr>
<td>Sulfasil</td>
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<td>BE/6000</td>
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The effects of SG on mRNA expression levels of inflammatory cytokines

To investigate the molecular mechanism of SG-induced wound healing activity, the expression levels of related inflammatory cytokines were examined. TNF-α, IL-1β, and IL-6 are the major genes that are generally involved in wound healing. On day 3, a significant decrease in the expression of TNF-α, IL-1β, and IL-6 in the wound tissue of SG/6000-treated rats compared with vehicle-treated rats was noted (Figure 5). Meanwhile,
SG/3000 and Sulfasil treatment showed modest effects.

Discussion

SG is a water-soluble natural product obtained from *Musa paradisiaca* plants. It has been documented that extracts of *Musa* species have antibacterial and antioxidant activities [25], and promote the healing of burn wounds [19]. Impaired wound healing is a common and potentially serious complication in diabetes [26]. In the present study, we used excisional wound models to investigate the role of SG in diabetic rats [27]. Furthermore, the wound healing effects of SG at a 1 : 6000 dilution were superior to those of SG at other concentrations in the initial stage of wound healing in STZ-induced diabetic rats. In contrast, the higher dilution of SG had better effectiveness in promoting wound healing in the late stage. However, CG did not promote wound-healing activity in STZ-induced diabetic rats. The findings of the present study indicated that SG/6000 was beneficial to skin wound healing in diabetic rats.

The wounds of diabetic rats in the present investigation showed prolonged inflammation, microvasculature aberrations that result in irreg-
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In the present study, SG/6000 treatment significantly decreased the number of inflammatory cytokines compared with that in other groups. Persistent inflammation in diabetes is associated with delayed wound healing. In diabetes, high glucose promotes inflammation through endothelial cell damage, the release of proinflammatory cytokines, and capillary permeability [35]. In the inflammatory phase, activated macrophages produce growth factors and inflammatory cytokines such as TNF-α, IL-1β, and IL-6 [34]. In the wound healing process, proinflammatory cytokines are involved in cell proliferation and differentiation as well [36]. IL-6 plays a crucial role in many wound healing phases, promoting the migration and proliferation of keratinocytes [37]. In the early stages of wound healing, TNF-α and IL-1β stimulate granulation tissue formation and neovascularization. TNF-α has been reported to regulate vascular endothelial growth factor levels and collagen synthesis in wound healing [38, 39]. IL-1β inhibits type I collagen synthesis during the repair phase of the wound healing response [40]. However, the persistence of inflammatory cytokines impairs the healing of diabetic wounds. SG treatment regulated these mediators by reducing proinflammatory cytokines at the early stage of wound healing and promoting wound repair at the late postinjury stages.

Our results indicated that SG/6000 has more potent anti-inflammatory wound healing stimulating properties than silver sulfadiazine. Silver sulfadiazine has been utilized in advanced wound care to improve the microenvironment of wounds [41]. The application of heparan sulfate can accelerate tissue regeneration in the early phase of tissue repair [42]. Nonetheless, a recent study indicated that silver dressings do not decrease the amount of bacteria in diabetic wounds [43]. A previous study documented that extracts of Musa paradisiaca have antibacterial activity against Pseudomonas aeruginosa and Staphylococcus aureus [44] and can increase the viability of collagen fibrils by increasing the strength of collagen fibers [45]. In addition, Musa species show antidiabetic and better ulcer healing effects in diabetic rats with co-occurring gastric ulcers [46]. Therefore, SG might be a target for the treatment of diabetes.

In conclusion, the present study showed that SG improves the healing process in diabetic rats through its anti-inflammatory effect and may help us cure wound injury in the clinic in the future. SG ointment has the potential to be a new alternative therapeutic strategy for wound healing.

Acknowledgments

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Conflict of interest

The authors declare no conflict of interest.

References


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