Crosstalk in human brain between globoid cell leucodystrophy and zinc-alpha-2-glycoprotein (ZAG), a biomarker of lipid catabolism

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Abstract

Zinc-alpha-2-glycoprotein (ZAG) is a protein identified as a lipid-mobilizing factor participating in a lipid catabolism. In spite of intensive studies conducted during last five decades, the role of this protein in processes of neurodegeneration remains unclear. The aim of our study was to examine the presence of ZAG protein in the brain of patients with Krabbe’s disease, which is considered as a psychosine lipidosis caused by a mutation of a known gene. We found intracellular and extracellular localization of ZAG in the brain of Krabbe’s disease patients but in the brain of control age-matched patients, ZAG was not detected. Distribution of ZAG in the brain suggests that the influx of ZAG into the brain involved a blood-brain barrier mechanism and adenoreceptors localized on astrocytes and some neurons.

Key words: Krabbe’s disease, globoid cell leucodystrophy, zinc-alpha-2-glycoprotein, cachexia, human brain, hypertrophied astrocytes, choroid plexus.

Introduction

The diagnosis of several neurodegenerative diseases is often difficult though it could be greatly enhanced by the use of disease-specific biomarkers [27]. In order to expand the search for such biomarkers, two-dimensional electrophoresis (2-DE) was previously used to separate proteins from different body fluids [6-8,26,35]. This method appeared useful not only for an effective identification of several proteins and their isoforms but also for an evaluation of quantitative and qualitative protein differences between the diseased and normal body state [11].

In the cerebrospinal fluid (CSF), a number of neuron-specific proteins and cytoskeleton markers have been found [21] and in the subsequent investigations, the important role of these proteins in the pathogenesis of some neurodegenerative processes was documented [1,3,28]. Zinc-alpha-2-glycoprotein (ZAG) was also separated by 2-DE, but in spite of intensive studies conducted during the last five decades the role of this protein in neurodegenerative processes remains unclear. ZAG is 41 kDa soluble protein first isolated from human plasma [4]. It was identified in different body fluids and organs, such as the liver, breast, lung, prostate and in both visceral and subcutaneous adipose tissue, suggesting its widespread exocrine function [2,17,30]. Finally, a role of ZAG was identified in patients with cancer...
cachexia as the lipid mobilizing factor associated with lipid catabolism of adipose tissue [13]. ZAG stimulates lipolysis through interaction with adrenoreceptors [24] and plays an important role in the function of brown adipose tissue, lipid catabolism and expenditure of energy from the body [13]. In the CSF, the level of ZAG seems to be connected precisely with pathological processes [11] but the presence of this protein in the brain during the process of neurodegeneration was not examined. The aim of the present study was to detect ZAG protein in the brain of patients with Krabbe’s disease that is the most common, severe, rapidly progressing neurodegeneration [9,19] and invariably fatal inherited metabolic disease caused by a known gene mutation [16,29,34].

Material and methods

Brain samples were obtained at autopsy of 11 patients with Krabbe’s disease (globoid cell leukodystrophy) who had died at 7-25 months of age. Tissue was fixed in formalin, embedded in paraffin, stained and clinical diagnosis was confirmed by neuropathological examination of cerebral sections in the Department of Developmental Neuropathology, Polish Academy of Sciences.

For the purpose of the present study, the brain paraffin blocks were drawn from the archive of the Medical Research Institute (Polish Academy of Sciences, Warsaw, Poland). The blocks containing samples of different brain regions were cut to 8 μm thick serial sections and were used for immunohistochemical study. Some sections were stained with cresyl violet for routine histological examination.

The same procedures of tissue preparation, immunohistochemical and morphological staining were used for the sections obtained from the brains of age-matched 11 control patients.

Immunohistochemistry

Immunohistochemistry was performed using the alkaline phosphatase-avidin-biotin conjugate or avidin-biotin-peroxidase complex system, according to the manufacturer’s recommendations (Vectastain Elite ABC Peroxidase Kit; Vector Laboratories).

Briefly, sections were dewaxed and hydrated through descending alcohols to water. For non-enzymatic antigen retrieval, sections were heated in 0.01 M sodium citrate buffer (pH 6.0) to 95°C and allowed to cool for 20 min at room temperature and washed with PBS. Then, sections were incubated in methanol/3% H2O2 solution for 20 min to quench endogenous peroxidase. Washed again in PBS and blocked with a solution containing PBS/5% normal serum of goat, rabbit or mouse for 2 h at room temperature, sections were incubated overnight at 4°C in solutions of primary antibodies (Table I).

In addition, glial cells were identified with: polyclonal glial fibrillary acidic protein (GFAP) antibodies generated against n-terminus (sc-6171) and c-terminus (sc-6170) domains of human GFAP origin (Santa Cruz) and Ferritin polyclonal antibody generated against human full length ferritin (DAKO).

Immunoreactions were visualized using biotinylated secondary antibodies and ABComplex/HRP or an alkaline phosphatase-avidin-biotin conjugate. Then, sections were lightly counterstained with Mayer’s haematoxylin.

For negative controls, primary antibodies were replaced with an appropriate isotypic normal mouse, rabbit or goat immunoglobulin fraction at matched protein concentration. These were included for the examination of each specimen and consistently produced negative results.

Table I. Antibodies generated against different domains of human zinc-alpha-2-glycoprotein (ZAG), source and dilution used in the study

<table>
<thead>
<tr>
<th>Antibody type</th>
<th>Antibody source</th>
<th>Catalogue</th>
<th>Antigen origin</th>
<th>Antigen domain</th>
<th>Dilution</th>
<th>Blocking peptide</th>
</tr>
</thead>
<tbody>
<tr>
<td>Monoclonal</td>
<td>Mouse</td>
<td>sc-21720</td>
<td>Human</td>
<td>nm</td>
<td>1:100</td>
<td>na</td>
</tr>
<tr>
<td>Monoclonal</td>
<td>Mouse</td>
<td>sc-13585</td>
<td>Human</td>
<td>fl</td>
<td>1:100</td>
<td>na</td>
</tr>
<tr>
<td>Monoclonal</td>
<td>Mouse</td>
<td>sc-21721</td>
<td>Human</td>
<td>nm</td>
<td>1:100</td>
<td>na</td>
</tr>
<tr>
<td>Polyclonal</td>
<td>Goat</td>
<td>sc-11238</td>
<td>Human</td>
<td>ir</td>
<td>1:50</td>
<td>sc-11238 P</td>
</tr>
</tbody>
</table>

sc – Santa Cruz, USA, na – not available, nm – not mentioned, fl – full length of ZAG, ir – internal region of ZAG
Results

The presence of ZAG protein was detected with a battery of different mono- and polyclonal antibodies generated against various fragments of human ZAG protein. The most precise localization of the antigen in human paraffin brain sections was found using a polyclonal antibody produced by goat against the internal region (ir) of ZAG protein. This antibody (sc-11238), as well as all other antibodies and chemicals used in the present study, were purchased from Santa Cruz Laboratories (USA).

In the brain of all patients affected by Krabbe’s disease, ZAG protein was found. The strong immunoreactivity documenting the presence of ZAG was localized precisely around capillaries of brain blood vessels. Such capillaries were scattered throughout the grey and white matter of brain hemispheres and cerebellum (Fig. 1). They were most numerous in the brain regions with a well-preserved structure of the nervous tissue. Blood vessels of a larger lumen diameter than capillaries contained strong immunoreactive blood serum but they were never surrounded by extracellular ZAG immunoreactivity (Fig. 2) as capillaries were. In brain regions where the white matter lesion was pronounced and where a spongy state developed, the presence of ZAG was detected in GFAP immunoreactive and hypertrophied astrocytes (Fig. 3). Intracellular localization of ZAG was found also in Purkinje cells and in all forms of their subsequent degeneration (Fig. 4). Ramified microglia as well as the small-sized and large-sized globoïd cells being immunoreactive for ferritin did not contain ZAG protein.

The presence of ZAG was found on epithelial cells of the choroid plexus of patients with Krabbe’s disease (Fig. 5) but not in control patients (Table II).

Discussion

Krabbe’s disease is an autosomal recessive disorder primarily affecting infants. In the great majority of cases, the symptoms appear during the first year of life. In a few cases, the symptoms come on during the first decade. Known mutations of the beta-galactosylcerebrosidase gene cause a dysfunction of lysosomal enzyme, which participates in the degradation of several glycosphingolipids [16,34], and lead to the accumulation of the galactosylsphingosine (psychosine) in the nervous system [14] and peripheral organs [15]. Thus, Krabbe’s disease is considered as a psychosine lipidosis.

Although psychosine is highly cytotoxic metabolite, which plays a critical role in the pathogenesis of Krabbe’s disease, there is no specific therapy available today.
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of the infantile form of Krabbe’s disease, the mechanism of its action in the pathophysiology of the disease has not yet been fully elucidated. One of the hypotheses suggests that psychosine mediates its toxic effect in cells by inhibition of the electron transport chain in mitochondria [10,31].

In such cells, the loss of mitochondrial membrane potential depends on the activation of the uncoupling protein (UCP) which expression, on the other hand, is mediated by zinc-alpha2-glycoprotein (ZAG) [25].

According to results of our present study, which documents for the first time that ZAG is present in the brain of patients with Krabbe’s disease, ZAG could be directly involved in the mechanism of psychosine neurotoxicity. Moreover, we observed that the main source of the brain ZAG was that circulating in blood plasma, which was strongly ZAG immuno-positive within the lumen of numerous blood vessels. However, only brain capillaries were surrounded by immuno-positive ZAG antigen suggesting that the influx transport of ZAG involves blood-brain barrier mechanisms that are localized on endothelium of these smallest brain blood vessels.

Fig. 2. ZAG detected in blood plasma of numerous brain blood vessels, in astrocytes but not in proliferated epithelial cells. Magn. ×100.

Fig. 3. Hypertrophic astrocytes containing ZAG. Brain areas undergoing spongy degeneration. Magn. ×200. Insert magn. ×400.
Intracellular localization of ZAG in hypertrophic astrocytes found in our study suggests also that ZAG protein as a lipid mobilizing factor could participate in the local lipid metabolism linked with the function of astrocytes in brain areas with myelin breakdown and/or in some types of nerve cells such as Purkinje cells of the cerebellum. It has been shown that both cell types (astrocytes and neurons) possess adrenergic receptors [12,22] that can bind ZAG [23,24]. Such local intracellular involvement of ZAG in the brain lipolysis could function as a process parallel to an independent neurotoxic effect of psychosine.

In addition, ZAG may serve in Krabbe’s disease as a marker of cachexia [33]. Several authors report-
ed that the cachectic state is an independent risk factor for mortality in different chronic diseases including cancer [18], chronic heart failure [5], chronic obstructive pulmonary disease [32] or rheumatoid arthritis [20].

In summary, results of our study document the presence of ZAG in the brain of Krabbe’s disease patients and suggest that this protein penetrates the brain through the blood-brain barrier.

Further studies are required to establish the association between the known inherited dysfunction of lysosomal enzyme and dysregulation of lipid metabolism in the brain of Krabbe’s disease patients and answer numerous questions concerning the understanding of both complex processes and whether ZAG levels in body fluids of these patients will precede known clinical symptoms of the disease.

References


