The transcription factor STAT3 is increasingly viewed as an oncogene. Activated STAT3 is pro-proliferative, antiapoptotic and oncogenic. There is growing evidence for the epigenetic regulation of STAT3 activity in cancer cells. We and others have previously reported that STAT3 activity in breast cancer and in lung endothelial and epithelial cells was inversely regulated by the tumor suppressor protein caveolin-1 (cav-1). In order to understand the mechanisms involved, we reexamined how STAT3 signaling, which is initiated in plasma membrane raft microdomains, traverses the cytoplasm to the nucleus. Using cellfractionation methods, we observed that by 15-30 min. after IL-6 treatment, up to two thirds of cytoplasmic Tyrphosphorylated STAT3 can be associated with the purified early endosome (EE) fraction (Rab-5-, EEA1-, TfR- and clathrin-positive fraction). Electron microscopy, immunofluorescence and detergent-dissection approaches confirmed the association of STAT3 and PY-STAT3 with early endosomes. STAT3transcriptional activation was inhibited by expression constructs for dominant negative dynamin K44A, epsin 2a, ampiphysin A1 and clathrin light chain, but enhanced by that for the active dynamin species MxA. Moreover, overexpression of the tumour suppressor cav-1 negatively regulated STAT3 signaling. Taken together, the data demonstrate strong epigenetic regulation of STAT3 signaling in cancer cells by virtue of dynamic membraneassociated trafficking along the caveolar (negative regulation) and endocytic (positive regulation) pathways.

**Key words:** STAT3, caveolin-1, oncogene, tumor suppressor, transcriptional regulation, signaling endosomes.

# **Epigenetic regulation of transcription factor STAT3 activity in cancer cells**

Epigenetyczna regulacja aktywności czynnika transkrypcyjnego STAT3 w komórkach nowotworowych

## Pravin B. Sehgal

Depts. of Cell Biology & Anatomy, and of Medicine, New York Medical College, New York, Valhalla, NY 10595, USA

## Introduction

The transcription factor STAT3 has been characterized as an oncogene in the context of breast cancer [1, 2]. Activated STAT3 is pro-proliferative, antiapoptotic and oncogenic [1, 2]. Several studies to investigate the relationship between nuclear STAT3 and phospho-STAT3 with breast cancer prognosis are currently ongoing [1-3]. The increased incidence of breast cancer with advancing age has been very recently linked to the increased secretion by breast stromal fibroblastic cells of interleukin-6 (IL-6)[4], which, in turn, activates STAT3 [5]. Conversely, inhibition of gp130 receptor signaling (the pathway used by IL-6-type cytokines) has been shown to reduce this "constitutive" activation of STAT3 in breast cancer cells and to reduce in vivo malignancy [6]. Moreover, mutations in the membrane raft and "tumor suppressor" protein caveolin-1 (cav-1) are common in human breast cancer (16% of cases) [reviewed in 7]. Experimentally, cav-1 loss (by genetic knockout) in the mouse leads to hyperactivation of STAT3 and STAT5, increased incidence of breast cancer and increased metastases of breast cancer to the lungs [7, 8]. STAT3 is also the major transcriptional factor activated in target cells by various gene therapy protocols which use the IL-6-type family of cytokines and their receptors. In this review we discuss a novel aspect of STAT3 function: the epigenetic regulation of its transcriptional activity by the tumor suppressor protein cav-1 and the trafficking of activated STAT3 through the cytoplasm along the membrane-associated endocytic pathway [9-11].

## IL-6 and STAT3 in breast cancer

In 1989 we were the first, using immunohistochemistry, to report the presence of IL-6 in neoplastic cell elements and stromal tissue of primary carcinomas of mammary, colonic, ovarian and endometrial origin as well as in adenocarcinomatous metastases to lymph nodes [12]. Additionally, we observed constitutive IL-6 immuno-staining in breast cancer cell lines (e.g. T47D cells) [12]. In vivo, IL-6 was an invariant presence in the host-tumor interaction, and is thought to contribute to symptoms of the paraneoplastic syndrome and cancer cachexia [reviewed in 13, 14].

In a line of research begun in the early 1990s, we observed that IL-6 increased breast ductal epithelial cell dyshesion and cell motility (in T47D and ZR-75-1 cells) [15-17]. Subcellular events accompanying this epithelioid to mesenchymal transformation included an IL-6-induced loss of focal adhesions and reduction in E-cadherin [15-18]. Other investigators also now relate IL-6, in cooperation with EGF, and scatter factor and TGF- $\alpha$ , to increased dyshesion and migration of breast cancer cells [reviewed in 17, 19, 20]. The increased incidence of breast cancer with advancing age has been very recently linked by Campisi et al. [4] to the increased secretion by breast

Czynnik transkrypcyjny STAT3 jest w coraz większym stopniu określany mianem onkogenu. Aktywowany STAT3 zwiększa proliferację komórek oraz ma działanie antyapoptotyczne i onkogenne. Coraz więcej dowodów wskazuje na epigenetyczną regulację STAT3 w komórkach nowotworowych. Poprzednio donoszono, że aktywność STAT3 w komórkach raka piersi oraz w komórkach śródbłonkowych i nabłonkowych płuc jest hamowana przez białko supresorowe kaweolinę-1 (cav-1). W celu zrozumienia mechanizmów zaangażowanych w ten proces regulacji ponownie zbadano, jak odbywa się przekazywanie sygnału za pośrednictwem STAT3 z błony komórkowej poprzez cytoplazmę do jądra. Używając metod frakcjonowania komórkowego zaobserwowano, że przez 15–30 min po poddaniu komórek działaniu interleukiny-6 (IL-6) nawet 2/3 fosforylowanego STAT3 może być związane z oczyszczoną wczesną frakcją endosomalną (EE – frakcja zawierająca Rab-5, EEA1, TfR i klatrynę). Mikroskopia elektronowa, immunofluorescencja i rozdzielanie w detergencie potwierdziły istnienie związania STAT3 i jego fosforylowanej postaci z wczesnymi endosomami. Aktywacja transkrypcji przez STAT3 była hamowana przy pomocy konstruktów ekspresyjnych wytwarzających negatywnie dominującą formę dynaminy K44A, epsynę 2a, ampifizynę A1 i lekki łańcuch klatryny, ale ulegała wzmocnieniu pod wpływem aktywnych postaci dynaminy MxA. Co więcej, nadekspresja supresora cav-1 negatywnie regulowała przekazywanie sygnału przez STAT3. Powyższe dane wskazują na silną epigenetyczną regulację przekazywania sygnału przez STAT3 w komórkach nowotworowych. Regulacja ta odbywa się zarówno poprzez wpływ na szlak kaweolarny (regulacja negatywna), jak i szlak endocytarny (regulacja pozytywna).

**Słowa kluczowe:** STAT3, kaweolina-1, onkogen, supresor guza, regulacja transkrypcji, endosomy sygnałowe. stromal fibroblastic cells of interleukin-6 (IL-6). Conversely, inhibition of gp130 receptor signaling (the pathway used by IL-6-type cytokines) has been shown to reduce this "constitutive" activation of STAT3 in breast cancer cells and to reduce in vivo malignancy [6]. The importance of STAT3 activation in breast cancer cells is further highlighted by (a) the recognition that scatter factor, a cytokine long investigated for its ability to promote breast cancer cell motility and "scattering" [17], is also now known to signal via the c-met receptor/STAT3 pathway [21], (b) that other growth factors and cytokines relevant to breast cancer invasiveness and metastasis (e.g. EGF, PDGF) also activate STAT3 signaling [5], (c) the identification of specific genes upregulated by STAT3 which contribute to enhanced breast cancer invasiveness (e.g. matrix metalloproteinase 9, MMP9) [1], and (d) the demonstration that small molecule inhibitors of activated STAT3 such as indirubin and STA-21 reduce the proliferative phenotype of breast cancer cells [22, 23].

# Initiation of cytokine/STAT3 signaling in plasma membrane raft microdomains

Diverse cytokines and growth factors, including IL-6, EGF, PDGF, scatter factor, activate the JAK/STAT3 signaling pathway leading to Tyrphosphorylation (by JAKs) and Ser-phosphorylation (via the ras-raf-MAPK pathway) of STAT3 [5]. We reported in 2002 that STAT3 activation at the plasma membrane took place in specialized microdomains called "rafts" (the "STATs in rafts" hypothesis; 24) in complexes with the raft protein cav-1 and the chaperones HSP90 and GRP58 [25, 26]. We have shown earlier that a consequence of reduced cav-1 in rafts is the hyperactivation of STAT3 signaling and, conversely, that overexpression of wt cav-1 inhibits STAT3 signaling in tumor cells (27, 28). Lisanti and colleagues have also shown hyperactivation of STAT5 in breast tissue of cav-1 KO mice together with breast epithelial cell hyperplasia [7, 29]. Moreover, breast tumor cells showed increased invasiveness and lung metastasis in cav-1 KO mice [29]. In additional studies, hyperactivation of STAT3 in different tissues has been confirmed in cav-1 KO mice [8]. Breast cancer cells expressing exogenously introduced wt cav-1 showed reduced tumorigenesis and invasiveness [29].

# Epigenetic regulation of STAT3 transcriptional activity along the endocytic pathway

Since 1999, research in this laboratory has resulted in novel insights into our understanding of how STAT3 works. Specifically, our previous position that latent STAT proteins do not exist in the cytosol as free monomers as per the original Darnell model of 1994 [reviewed in 5] but in large complexes [9, 30, 31] is now widely accepted, including by the Darnell group [32, 33]. In recent years we [11] and Bild et al. [10] have discovered a novel aspect of STAT3 trafficking and activation in the cytoplasm: the majority (approximately 75%) of cytoplasmic trafficking of PY-STAT3 is membrane associated (Fig. 1). Using cell-fractionation methods, we observed that approximately 5% of cytoplasmic STAT3 was constitutively associated with the purified early endosome (EE) fraction in human hepatoma Hep3B cells (which have low endogenous cav-1). By 15-30 min. after IL-6 treatment, up to two thirds of cytoplasmic Tyr-phosphorylated STAT3 can be associated with the purified early endosome fraction (Rab-5-, EEA1-, TfR- and clathrin-positive fraction). Electron microscopy, immunofluorescence and detergent-dissection approaches confirmed the association of STAT3 and PY-STAT3 with early endosomes. Further immunofluorescence and cell fractionation studies confirmed the association of STAT3 with the Rab7 and LAMP1, 2-positive late endosomal/lysosomal compartment. Biochemically, STAT3 was constitutively in complexes with clathrin heavy chain (CHC) in membrane and cytosolic compartments. As with clathrin, the membrane association of STAT3 was dynamic in that within 15 min. of treatment with the vicinal-thiol crosslinker phenylarsine oxide (PAO) there was a dramatic increase in bulk STAT3

association with sedimentable membranes. The functional contribution of PY-STAT3 association with the endocytic pathway was evaluated in transient transfection assays using IL-6-inducible STAT3-reporter-luciferase constructs and selective regulators of this pathway. STAT3transcriptional activation was inhibited by expression constructs for dominant negative dynamin K44A, epsin 2a, ampiphysin A1 and clathrin light chain, but enhanced by that for the active dynamin species MxA. Taken together, the new data [11 and citations therein concerning the negative STAT3 regulators hepatocyte growth factor receptor tyrosine kinase substrate HRS and schwanomin] demonstrate strong epigenetic regulation of STAT3 signaling by virtue of dynamic membrane-associated trafficking along the caveolar (negative regulation) and endocytic (positive regulation) pathways. In other words, STAT3 dynamics at the plasma membrane and in the cytoplasm are inextricably linked to those of caveolin, clathrin, endocytosis and the biochemical regulatory machineries involved at intracellular membrane surfaces (the "STAT3 signaling endosome hypothesis"; Fig. 1).

# The "STAT3 signaling endosome hypothesis": a platform for cross-talk with other pathways

As a template for understanding where our studies fit into the broader signaling literature, the convergence of caveolar and endocytic pathways in the regulation of cellular signaling is becoming apparent in several other experimental systems. Available data show convincingly the merging of caveolar and endosomal pathways [34-36]. With respect to ligandactivated signaling, the formulation of a "signaling endosome hypothesis" in the mid 1990s with respect to NGF action has drawn attention to the ability of clathrin-coated endosomal vesicles to mediate rapid and directional transit of signaling molecules through the cytoplasm in [37-39]. In PC12 cells, NGF recruited CHC and TrkA into endosomal membraneassociated complexes with AP2 adaptor which then served as a platform for the ras-MAPK pathway [37-39]. Moreover, this endomembrane-associated signaling pathway was bifurcated in that of the two NGF receptors p75NTR was eventually partitioned towards recycling endosomes while TrkA was directed towards late endosomes/lysosomes for degradation [39]. Additional evidence for a functional bifurcation during internalization comes from the demonstration that ligand-induced raft/TGF-BR/Smad transcriptional signaling was regulated negatively by cav-1 as a result of targeting to lysosomes and regulated positively by targeting to endosomes [40]. The complexities of dual positive and negative signaling during vesicular internalization have been further highlighted by the observation that clathrin-dependent and clathrinindependent pathways can be differentially engaged in the case of EGF signaling based upon ligand concentration and ubiguitination of epsin/AP2 adaptors [41, 42].



# STAT3 signaling endosome hypothesis

Breast cancer: loss/mutations in cav-1 >hyperactivated STAT3>? mehanisms

**Fig. 1.** Epigenetic regulation of the transcriptional activity of STAT3: regulation by caveolin-1 and clathrin *Ryc. 1.* Epigenetyczna regulacja aktywności STAT3 jako czynnika transkrypcyjnego: regulacja przez kaweolinę-1 I klatrynę

In the case of IL-6/gp130/STAT3 signaling, it is clear that there is constitutive internalization of gp130 in interaction with the AP2 adaptor complex [43]. Moreover, Bild and colleagues have shown that EGF- and PDGF-activated STAT3 co-localize in the cytoplasm with vesicular elements which also contain the  $\alpha$ -adaptin polypeptide of the AP2 complex [10]. More generally, receptors for diverse cytokines which activate STAT signaling are known to traffic along the endocytic pathway in a manner dependent upon their Leu-Leu or Leu-Ile internalization motif [44]. Recently Meads and Medveczky [45] have interpreted their findings on the occurrence of gp130/STAT3 signaling, as assayed in terms of Tyr-phosphorylation of cytoplasmic STAT3, by vIL-6 mutants which do not appear to be secreted, as activation from within the endoplasmic reticulum/Golgi compartments. Similar observations of activation of the JAK/STAT pathway by secretion defective truncated interferons  $\alpha$ ,  $\beta$  or  $\gamma$  apparently from within the cell have also been reported [46 and citations therein]. Our hypothesis concerning the constitutive and dynamic association of STATs with cytoplasmic vesicular elements provides a basis for understanding these observations.

Is there a kinetic advantage of the endosomal signaling pathway compared to a cytosolic random walk mechanism? Following up on the original proposal of a "signaling endosome" with NGF, Howe [47] carried out computer modeling studies of PY-STAT3 traverse from the plasma membrane to the nucleus using a non-directional random walk or a directional endosomal trafficking paradigm. Even in 2-D simulations using conservative assumptions of a Stokes radius for STAT3 equivalent to 100 kDa, cellular geometry and cytosolic viscosity, directional endosomal trafficking reached the nucleus from the plasma membrane in 1 sec., but the random walk from the plasma membrane failed to do so. Thus, directional endosomal trafficking was kinetically more efficient than a random walk through the free cytosol [47].

## Conclusions

The space between the plasma membrane and the cell nucleus is not "empty" as is depicted in innumerable schemes describing signal transduction initiated by cytokines and growth factors. The trans-cytoplasmic traverse of an activation signaling appears to be efficiently mediated in association with the cytoplasmic membrane pathways – the endocytic pathway – which exerts both positive and negative influence on the eventual nuclear targeting and function of the respective activated transcription factors such as STAT3.

## Acknowledgements

Research in the author's laboratory was supported by Research Grant HL-077031 from the National Institutes of Health.

### References

1. Dechow T, Bromberg, J. 2003. Constitutively active STATs and cellular transformation. In: Signal Transducers and Activators of

Transcription: Activation and Biology, Sehgal PB, Levy DE, Hirano T (eds), Kluwer Academic Press, Dordrecht, The Netherlands, 2003; 637-644.

- Buettner R, Kortylewski M, Pardoff D, Yu H, Jove R. STAT proteins as molecular targets for cancer therapy, In: Signal Transducers and Activators of Transcription: Activation and Biology, Sehgal PB, Levy DE, Hirano T (eds), Kluwer Academic Press, Dordrecht, The Netherlands, 2003; 645-661.
- Dolled-Filhart M, Rimm DL, Jaks and Stats as biomarkers of disease. In: Signal Transducers and Activators of Transcription: Activation and Biology, Sehgal PB, Levy DE, Hirano T (eds), Kluwer Academic Press, Dordrecht, The Netherlands, 2003; 697-720.
- 4. Campisi J. Cellular snescence, cancer and aging. Mol Biol. Cell 2004; 15: 354a (and Symposium Lecture at the Annual Meeting of the Am. Soc. Cell Biology in Washington D. C., Dec 4-8, 2004).
- 5. Sehgal PB, Levy DE, Hirano T. Signal Transducers and Activators of Transcription: Activation and Biology, Kluwer Academic Press, Dordrecht, The Netherlands, 2003; 1-746.
- Selander KS, Li L, Watson L, Merrell M, Dahmen H, Heinrich PC, Muller-Newen G, Harris KW. Inhibition of gp130 signaling in breast cancer blocks constitutive activation of Stat3 and inhibits in vivo malignancy. Cencer Res 2004; 64: 6924-6933.
- Park DS, Lee H, Frank PG, et al. Caveolin-1-deficient mice show accelerated mammary gland development during pregnancy, premature lactation, and hyperactivation of the Jak-2/STAT5a signaling cascade. Mol Biol Cell 2002; 13: 3416-3430.
- Jasmin JF, Mercier L, Hnasko R, Cheung MW, Tanowitz HB, Dupuis J, Lisanti, MP. Lung remodeling and pulmonary hypertension after myocardial infarction: pathogenic role of reduced caveolin expression. Cardiovasc Res 2004; 63: 747-755.
- Sehgal PB. STAT signaling through the cytoplasmic compartment: consideration of a new paradigm. Cell Signaling 2000; 12: 525-535.
- Bild AH, Turkson J, Jove R. Cytoplasmic transport of Stat3 by receptormediated endocytosis. EMBO J. 2000; 21: 3255-3263.
- 11. Shah M, Patel K, Mukhopadhyay S, Xu F, Guo G, Sehgal PB. Membrane-associated STAT3 and PY-STAT3 in the cytoplasm. J Biol Chem 2006; 281: 7302-7308.
- 12. Tabibzadeh SS, Poubouridis D, May LT, Sehgal PB. Interleukin-6 immunoreactivity in human tumors. Am J Path 1989; 135: 427-433.
- 13. Sehgal PB. Interleukin-6 in infection and cancer. Proc Soc Exp Biol Med 1990; 195: 183-191.
- 14. Sehgal PB. Cytokines in the host-tumor interaction. In: Molecular Aspects of Cancer and Its Therapy, Mackiewicz A, Sehgal PB (Eds), MCBU Series, Birkhauser Verlag, Basel, 1998; 89-106.
- Tamm I, Cardinale I, Krueger J, Murphy JS, May LT, Sehgal PB. Interleukin-6 decreases cell-cell association and increases motility of ductal breast carcinoma cells. J Exp Med 1989; 170: 1649-1669.
- 16. Tamm I., Cardinale I, Sehgal PB. Interleukin-6 and 12-O-tetradecanoyl phosbol-13-acetate act synergistically in inducing cell-cell separation and migration of human breast carcinoma cells. Cytokine 1991; 3: 212-223.
- 17. Sehgal PB, Tamm I. Interleukin-6 enhances motility of breast carcinoma cells. In: Cell Motility Factors, Goldberg ID, Rosen E (Eds), Birkhauser-Verlag, Basel, 1991; 178-193.
- Tamm I, Cardinale I, Kikuchi T, Krueger JG. E-cadherin distribution in interleukin-6-induced cell-cell separation of ductal breast carcinoma cells. Proc Natl Acad Sci USA 1994; 91: 4338-4342.
- Tamm I, Kikuchi T, Cardinale I, Murphy JS, Krueger JG. Cytokines in breast cancer cell dyshesion. In: Molecular Aspects of Cancer and Its Therapy, . Mackiewicz A, Sehgal PB, MCBU Series, Birkhauser Verlag, Basel, 1998; 21-43.
- 20. Badache A, Hynes NE. Interleukin-6 inhibits proliferation and, in cooperation with an epidermal growth factor receptor autocrine loop, increases migration of T47D breast cancer cells. Cancer Res 2001; 61: 383-391.
- Cramer A, Kleiner S, Westermann M, Meissner A, Lange A, Friedrich K. Activation of the c-Met receptor complex in fibroblasts drives invasive cell behaviour by signaling through transcription factor STAT3. J Cell Biochem 2005; 95: 805-816.
- 22. Nam S, Buettner R, Turkson J, et al. Indirubin derivatives inhibit Stat3 signaling and induce apoptosis in human cancer cells. Proc Natl Acad Sci USA 2005; 102: 5998-6003.

- 23. Song H, Wang R, Wang S, Lin J. A low-molecular weight compound discovered through virtual database screening inhibits Stat3 function in breast cancer cells. Proc Natl Acad Sci USA 2005; 102: 4700-4705.
- 24. Sehgal PB, Guo GG, Shah M, Kumar V, Patel K. Cytokine signaling: STATs in plasma membrane rafts. J Biol Chem. 2002; 277: 12067-12074.
- 25. Shah M, Patel K, Fried VA, Sehgal PB. Interactions of STAT3 with caveolin-1 and heat shock protein 90 in plasma membrane rafts and cytosolic complexes: preservation of cytokine signaling during fever. J Biol Chem 2002; 277: 45662-45669.
- 26. Sehgal PB. Plasma membrane rafts and chaperones in cytokine/STAT signaling. Acta Biochemica Polonica 2003; 50: 583-594
- 27. Mathew R, Huang J, Shah M, Patel K, Gewitz M, Sehgal PB.Disruption of endothelial-cell caveolin- $1\alpha$ /raft scaffolding during development of monocrotaline-induced pulmonary hypertension. Circulation 2004; 110: 1499-1506.
- 28. Shah M, Patel K, Sehgal PB. Monocrotaline pyrrole-induced endothelial cell megalocytosis involves a Golgi-blockade mechanism. Am J Physiol Cell. Physiol 2005; 288: C850-C862.
- 29. Williams TM, Medina F, Badano I, et al. Caveolin-1 gene disruption promotes mammary tumorigenesis and dramatically enhances lung metastasisin vivo. Role of cav-1 in cell invasiveness and matrix metalloproteinase (MMP-2/9) secretion. J Biol Chem 2004; 279: 51630-51640.
- Ndubuisi MI, Guo GG., Fried VA, Etlinger JD, Sehgal PB. Cellular physiology of STAT3: where's the cytoplasmic monomer? J Biol Chem 199; 274: 25499-25509.
- 31. Guo GG, Patel K, Kumar V, Shah M, Fried VA, Etlinger JD, Sehgal, P.B. Association of the chaperone glucose-regulated protein 58 (GRP58/ER-60/ERp57) with Stat3 in the cytosol and plasma membrane complexes. J Interferon & Cyt Res. 2002; 22: 555-563.
- Mao X, Ren Z, Parker GN, et al. Structural bases of unphosphorylated STAT1 association and receptor binding. Mol Cell 2005; 17: 761-771.
- 33. Zhong M, Henriksen MA, Takeuchi, K, et al. Implications of an antiparallel dimeric structure of nonphosphorylated STAT1 for the activation-inactivation cycle. Proc Natl Acad Sci USA 2005; 102: 3966-3971.
- 34. Parton RG. Caveolae meet endosomes: a stable relationship? Developmental Cell 2004; 7: 458-460.
- 35. Sadir R, Lambert A, Lortat-Jacob H, Morel G. Caveolae and clathrincoated vesicles: two possible internalization pathways for IFNgamma and IFN-gamma receptors. Cytokine 2001; 14: 19-26.
- 36. Sharma DK, Choudhury A, Singh RD, Wheatley CL, Marks DL, Pagano RE.. Glycosphingolipids internalized via caveolar-related endocytosis rapidly merge with clathrin pathway in early endosomes and form microdomains for recycling. J Biol Chem 2003; 278: 7564-7572.
- Howe CL, Valletta JS, Rusnak AS, Mobley WC. NGF signaling from clathrin-coated vesicles: evidence that signaling endosomes serve as a platform for the Ras-MAPK pathway. Neuron 2001; 32: 801-814.
- 38. Leof EB. Growth factor receptor signalling: location, location, location. Trends Cell Biol 2000; 10: 343-348.
- Saxena S, Howe CL, Cosgaya JM, et al. Differential endocytic sorting of p75NTR and TrkA in response to NGF: a role for late endosomes in TrkA trafficking. Mol Cell Neurosci. 2005; 28: 571-587.
- 40. DiGuglielmo GM, LeRoy C, Goodfellow AF, Wrana JL. Distinct endocytic pathways regulate TGFβ receptor signalling and turnover. Nature Cell Biol 2003; 5: 410-421.
- 41. Chen H, DeCamilli P. The association of epsin with ubiquitinated cargo along the endocytic pathway is negatively regulated by its interaction with clathrin. Proc Natl Acad Sci USA 2005; 102: 2766-2771.
- 42. Sigismund S, Woelk T, Puri C, et al. Clathrin-independent endocytosis of ubiquitinated cargos. Proc Natl Acad Sci USA 2005; 102: 2760-2765.
- 43. Thiel S, Dahmen H, Martens A, Muller-Newen G, Schaper F, Heinrich PC, Graeve L. Constitutive internalization and association with adaptor protein-2 of the interleukin-6 signal transducer gp130. FEBS Lett 1998; 441: 231-234.
- 44. Theil S, Behrmann I, Timmermann A, et al. Identification of a Leu-Ile internalization motif within the cytoplasmic domain of the leukemia inhibitory factor receptor. Biochem J 1999; 339: 15-19.

- 45. Meads MB, Medveczky PG. Kaposi's sarcoma-associated herpesvirus-encoded viral interleukin-6 is secreted and modified differently than human interleukin-6: evidence for a unique autocrine signaling mechanism. J Biol Chem 2004; 279: 51793-51803.
- 46. Shin-Ya M, Hirai H, Satoh E, et al. Intracellular interferon triggers Jak/STAT signaling cascade and induces p53 dependent antiviral protection. Biochem Biophys Res Commun 2005; 329: 1139-1146.
- 47. Howe CL Modelling the signaling endosome hypothesis: why a drive to the nucleus is better than a (random) walk. Theor Biol Med Model 2005; 2: 43.

### Address for correspondence

### Dr. Pravin B. Sehgal

Rm. 201 Basic Sciences Building New York Medical College Valhalla, NY 10595 Tel: 914-594-4196 Fax: 914-594-4825 E-mail: Pravin\_sehgal@nymc.edu