Modulation of immune functions by oestrogens. Part I

Immunomodulacyjna rola estrogenów. Część I

Jacek R. Wilczyński^{1, 2}

¹Department of Gynecological Surgery, "Polish Mother's Memorial Hospital" Research Institute, Lodz, Poland Head: prof. dr hab. n. med. Marian Szpakowski

²Deptartment of Gynecology, Chair of Obstetrics & Surgical Gynecology, Medical University of Lodz, Poland Head: prof. dr hab. n. med. Jacek R. Wilczyński

Przegląd Menopauzalny 2010; 2: 59-62

Summary

One of the most important functions of oestrogens is modulation of the immune system. By interactions with specific oestrogen receptors $ER\alpha$ and $ER\beta$ these sex steroid hormones are capable of regulating many aspects of both natural and adaptive immunity. Aging and the hypoestrogenic state can therefore influence immunological functions. Use of drugs belonging to SERM or isoflavones could also modify the immune response in post-menopausal women. All these phenomena might influence the risk of cancer initiation and the natural course of tumour growth.

Key words: immunology, oestrogens.

Streszczenie

Jedną z najważniejszych funkcji estrogenów jest zdolność modulowania działania układu immunologicznego. Poprzez interakcje ze swoistymi receptorami estrogenowymi ERα i ERβ estrogeny są zdolne do regulowania zarówno elementów odporności naturalnej, jak i swoistej. Dlatego starzenie się i towarzyszący mu stan hipoestrogenizmu mogą wpływać na funkcje immunologiczne. Podobnie zastosowanie u kobiet po menopauzie leków z grup SERM i izoflawonów może modyfikować odpowiedź immunologiczną. Zjawiska powyższe mogłyby stanowić mechanizm wpływający zarówno na ryzyko powstania, jak i na charakter wzrostu nowotworów.

Słowa kluczowe: immunologia, estrogeny.

Introduction

It is a commonly accepted fact that the biological functions of oestrogens are not limited to breast and reproductive organs, but that these hormones also influence body metabolism and function of the cardio-vascular system, brain and bones. However, one of the most important beneficiaries of oestrogen regulatory capabilities is the immune system. The biological action of oestrogens is mediated through specific high-affinity oestrogen receptors (ER α and ER β) which are ligand-dependent transcription factors [1, 2]. The genes subjected to oestrogen regulation include those responsible for cell differentiation, growth and survival [3]. The interaction of oestrogens with ER can occur directly, through oestrogen response elements (EREs), or indirectly through mediation of other transcription factors such as activator

protein 1 (AP1), factor SP1 or nuclear factor κB (NF- κB) [4]. The receptor ER β activates the same genes in the response to oestradiol stimulation; however, its efficacy is less than that of ER α . Generally, ER β plays the role of an inhibitor of $ER\alpha$ in cells where both types of ER are present [5]. The final effect of oestrogen depends on its concentration, the relative cellular density of both ER, and co-activation by different factors [6]. Besides, oestrogens show probably ER-independent abilities of direct modulation of intracellular signalling pathways, through stimulation of kinases, phosphatases, and calcium channels [7]. At least one of the ER is expressed inside lymphoid organs, on the surface of haematopoietic progenitors in bone marrow, as well as on peripheral lymphocytes, natural killer cells (NK), neutrophils, dendritic cells (DC), monocytes and macrophages [8-13].

Address for correspondence:

prof. dr hab. n. med. Jacek R. Wilczyński, Deptartment of Gynecology, Chair of Obstetrics & Surgical Gynecology, Medical University of Lodz, 4 Kosciuszki Str., 90-419 Lodz, Poland

Influence of oestrogens on immune function

Observations of immune functions in both females and males indicate the existence of gender-based differences. Compared to men, women have increased immunity towards infections caused by both extra- and intracellular pathogens, and generate stronger responses after vaccinations, but unfortunately show a predisposition to autoimmune reactions [14-16]. However, during the reproductive period women have better control over excessive inflammation, and thus are less prone to septic complications [17, 18]. Those gender-based differences indicated that sex steroid hormones, including oestrogens, could modify the immune response. The results of many studies performed on both mice and humans show that oestrogens enhance especially the humoral response, but the influence of cellular immunity seems to be more complex, and is either stimulatory or suppressive dependent on dose of oestrogen, duration of action and immune cell population [19-22].

Oestrogens and innate immunity

Animal studies clearly indicate that oestrogens can modulate the immune response in the reproductive tract of studied mice. The concentration of molecules of known antimicrobial activity present in cervical and uterine fluids such as lactoferrin and lysozyme is regulated by oestradiol concentrations [23, 24]. Oestradiol administered into mice caused an increase in uterine vascular permeability, which resulted in an increase of extravasated C1q, C2, C3, C4, and C6 components of the complement system in uterine tissues and luminal fluid [25]. During oestrus, characterized by the highest concentration of oestrogens, recruitment of neutrophils, macrophages and DC into the endometrium and uterine stroma was significantly increased [26-28]. Ovariectomized animals did not show such changes, unless they were given hormonal supplementation [29]. The number and function of uterine NK cells infiltrating the endometrium are also oestradiol dependent [30]. Exposure to oestrogens enhances cytotoxicity of peripheral blood NK cells and increases IFNy secretion [31, 32]. Human studies support these results, and indicate that postmenopausal women show decreased NK activity, which returns to the premenopausal level during hormonal therapy [33]. Oestrogens indicate functional ambiguity towards innate immunity, as depending on their concentration and time of action, they are capable of down-regulation of an exaggerated immune response inside genital tracts by reduction of neutrophil infiltration and decrease of tumour necrosis factor- α (TNF α) production [34-36].

Oestrogens are also modulators of DC function, decreasing their migration, maturation and capaci-

ty for antigen presentation. Mouse studies indicated that oestradiol negatively regulated secretion of $TNF\alpha$, interferon- γ (IFN γ) and interleukin (IL)-12 by DC, thus shifting cytokine production towards the Th2 profile (IL-4, IL-10) [37]. Inside reproductive organs oestrogens were able to disturb antigen presentation of both professional antigen-presenting cells (APC), such as DC and macrophages, and of epithelial cells functioning as non-classical APC [38, 39]. The above-mentioned mechanisms seem to be regulated by oestrogen-dependent secretion of transforming growth factor- β (TGF β), inhibitory cytokine produced by immature DC [40, 41]. Oestrogens also promote DC survival, as administration of oestradiol immediately after trauma-haemorrhage prevented splenic DC apoptosis and dysfunction observed normally in these cases [42]. However, some investigations have put more criticism into our understanding of oestrogen-regulated DC function, because it was found that the regulatory capacity of oestrogens depends on their dose and time of action. Supra-physiological doses of oestradiol caused opposite effects than those described above. A short time exposure to higher oestradiol doses stimulated secretion of pro-inflammatory Th1 cytokines, while a long time exposure increased splenic DC antigen presentation capacity [43, 44]. Oestrogens could also have different influence on particular DC subpopulations (Langerhans cells, myeloid, splenic and IFN-producing killer DC) [45, 46], and exert opposing effects on DC bone marrow differentiation mediated by granulocyte-macrophage colony stimulating factor (GM-CSF) and Flt3 ligand [47]. As the Flt3-mediated pathway is used mainly in normal circumstances, while the GM-CSF-mediated pathway is used during inflammation, oestrogens seem to affect DC differentiation dependent on the status of homeostasis or pathology, as well as on cytokine milieu [48].

The influence of oestrogens on macrophage function consists of enhancement of expression of inducible nitric oxide synthase (iNOS) and proinflammatory IL-1, IL-6, IFN_γ and TNF_α cytokines [49, 50]. This oestrogen action was also confirmed in a mouse model of encephalitis. In this model microglial cells (macrophage homologues) of ovariectomized mice were unable to produce TNF α and IL-12 during brain infection [51]. However, as in the case of DC, the precise oestrogen action depends on its dose and differentiation state of monocytes and macrophages [52, 53]. Oestrogen induced Fas/FasL-dependent apoptosis in monocytes expressing $ER\beta$, but not in macrophages expressing $ER\alpha$ [54]. It was found in "in vitro" studies that low concentrations of oestrogen induced iNOS activity in resting macrophages, but at higher concentrations oestrogen appeared to reduce iNOS activity [54-56]. According to some authors [57] even in the presence of physiological oestradiol concentrations activation of macrophages by lipopolysaccharide (LPS) was ineffective, due to inhibitory effects of oestrogen on NK- κ B function, and induction of macrophage apoptosis. These observations support the thesis of antiinflammatory oestrogen function in high/normal dose and suggest that it affects the activation of macrophages by toll-like receptor (TLR) ligands [58].

References

- McKenna NJ, O'Malley BW. Nuclear receptors, coregulators, ligands, and selective receptor modulators: making sense of the patchwork quilt. Ann N Y Acad Sci 2001; 949: 3-5.
- Means AR, Comstock JP, Rosenfeld GC, et al. Ovalbumin messenger RNA of chick oviduct: partial characterization, estrogen dependence, and translation in vitro. Proc Natl Acad Sci USA 1972; 69: 1146-50.
- O'Lone R, Frith MC, Karlsson EK, et al. Genomic targets of nuclear estrogen receptors. Mol Endocrinol 2004; 18: 1859-75.
- 4. McKay LI, Cidlowski JA. Molecular Control of Immune/Inflammatory Responses: Interactions Between Nuclear Factor-kappa B and Steroid Receptor-Signaling Pathways. Endocrine Reviews 1999; 20: 435-59.
- Hall JM, McDonnell DP. The estrogen receptor beta-isoform (ERbeta) of the human estrogen receptor modulates ERalpha transcriptional activity and is a key regulator of the cellular response to estrogens and antiestrogens. Endocrinology 1999; 140: 5566-78.
- McDonnell DP. The molecular determinants of estrogen receptor pharmacology. Maturitas 2004; 48: S7-S12.
- Heldring N, Pike A, Andersson S, et al. Estrogen receptors: how do they signal and what are their targets. Physiol Rev 2007; 87: 905-31.
- Phiel KL, Henderson RA, Adelman SJ, et al. Differential estrogen receptor gene expression in human peripheral blood mononuclear cell populations. Immunol Lett 2005; 97: 107-13.
- Igarashi H, Kouro T, Yokota T, et al. Age and stage dependency of estrogen receptor expression by lymphocyte precursors. Proc Natl Acad Sci USA 2001; 98: 15131-6.
- Komi J, Lassila O. Nonsteroidal anti-estrogens inhibit the functional differentiation of human monocyte-derived dendritic cells. Blood 2000; 95: 2875-82.
- Harkonen PL, Vaananen HK. Monocyte-macrophage system as a target for estrogen and selective estrogen receptor modulators. Ann NY Acad Sci 2006; 1089: 218-27.
- Mor G, Sapi E, Abrahams VM, et al. Interaction of the estrogen receptors with the Fas ligand promoter in human monocytes. J Immunol 2003; 170: 114-22.
- 13. Grimaldi CM, Cleary J, Dagtas AS, et al. Estrogen alters thresholds for B cell apoptosis and activation. J Clin Invest 2002; 109: 1625-33.
- Marriott I, Huet-Hudson YM. Sexual dimorphism in innate immune responses to infectious organisms. Immunol Res 2006: 34: 177-92.
- 15. Whitacre CC. Sex differences in autoimmune disease. Nat Immunol 2001; 2: 777-80.
- Cutolo M, Straub RH. Insights into endocrine-immunological disturbances in autoimmunity and their impact on treatment. Arth Res Ther 2009; 11: 218-25.
- Kovacs EJ. Aging, traumatic injury, and estrogen treatment. Exp Gerontol 2005; 40: 549-55.
- 18. Straub RH. The complex role of estrogens in inflammation. Endocr Rev 2007; 28: 521-74.
- 19. Suzuki S, Brown CM, Dela Cruz CD, et al. Timing of estrogen therapy after ovariectomy dictates the efficacy of its neuroprotective and antiinflammatory actions. Proc Natl Acad Sci USA 2007; 104: 6013-8.
- Naugler WET, Sakurai S, Kim S, et al. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. Science 2007; 317: 121-4.
- 21. Beagley KW, Gockel CM. Regulation of innate and adaptive immunity by the female sex hormones oestradiol and progesterone. FEMS Immunol Med Microbiol 2003; 38: 13-22.
- 22. Verthelyi D. Female's heightened immune status: estrogen, T cells, and inducible nitric oxide synthase in the balance. Endocrinology 2006; 147: 659-61.
- 23. Cohen MS, Britigan BE, French M, et al. Preliminary observations on lactoferrin secretion in human vaginal mucus: variation during the menstrual

cycle, evidence of hormonal regulation, and implications for infection with Neisseria gonorrhoeae. Am J Obstet Gynecol 1987; 157: 1122-5.

- Walmer DK, Wrona MA, Hughes CL, et al. Lactoferrin expression in the mouse reproductive tract during the natural estrous cycle: correlation with circulating estradiol and progesterone. Endocrinology 1992; 131: 1458-66.
- 25. Rhen T, Cidlowski JA. Estrogens and Glucocorticoids Have Opposing Effects on the Amount and Latent Activity of Complement Proteins in the Rat Uterus. Biol Reprod 2006; 74: 265-74.
- Sonoda Y, Mukaida N, Wang JB, et al. Physiologic regulation of postovulatory neutrophil migration into vagina in mice by a C-X-C chemokine-(s). J Immunol 1998; 160: 6159-65.
- 27. Penny LA. Monocyte chemoattractant protein 1 in luteolysis. Rev Reprod 2000; 5: 63-6.
- Reibiger I, Spanel-Borowski K. Difference in localization of eosinophils and mast cells in the bovine ovary. J Reprod Fertil 2000; 118: 243-9.
- De M, Wood GW. Influence of oestrogen and progesterone on macrophage distribution in the mouse uterus. J Endocrinol 1990; 126: 417-24.
- Henderson TA, Saunders PT, Moffet-King A, et al. Steroid receptor expression in uterine natural killer cells. J Clin Endocrinol Metab 2003; 88: 440-449.
- Nakaya M, Tachibana H, Hamada K. Effect of estrogens on the interferon-γ producing cell population of mouse splenocytes. Biosci Biotechnol Biochem 2006; 70: 47-53.
- 32. Sorachi K, Kumagai S, Sugita J, et al. Enhancing effect of 17 β -estradiol on human NK cell activity. Immunol Lett 1993; 36: 31-5.
- 33. Yang JH, Chen CD, Wu MY, et al. Hormone replacement therapy reverses the decrease in natural killer cytotoxicity but does not reverse the decreases in the T-cell subpopulation or interferon-gamma production in postmenopausal women. Fertil Steril 2000; 74: 261-7.
- Cuzzocrea S, Mazzon E, Sautebin L, et al. The protective role of endogenous estrogens in carrageenan-induced lung injury in the rat. Mol Med 2001; 7: 478-87.
- Ozveri ES, Bozkurt A, Haklar G, et al. Estrogens ameliorate remote organ inflammation induced by burn injury in rats. Inflamm Res 2001; 50: 585-91.
- 36. Eckhoff DE, Bilbao G, Frenette L, et al. 17-Beta-estradiol protects the liver against warm ischemia/reperfusion injury and is associated with increased serum nitric oxide and decreased tumor necrosis factor-alpha. Surgery 2002; 132: 302-9.
- Liu HY, Buenafe AC, Matejuk A, et al. Estrogen inhibition of EAE involves effects on dendritic cell function. J Neurosci Res 2002; 70: 238-48.
- Prabhala RH, Wira CR. Sex hormone and IL-6 regulation of antigen presentation in the female reproductive tract mucosal tissues. J Immunol 1995; 155: 5566-73.
- Wira CR, Rossoll RM. Antigen-presenting cells in the female reproductive tract: influence of sex hormones on antigen presentation in the vagina. Immunology 1995; 84: 505-8.
- Geissmann F, Revy P, Regnault A, et al. TGF-beta 1 prevents the noncognate maturation of human dendritic Langerhans cells. J Immunol 1999; 162: 4567-75.
- Wira CR, Roche MA, Rossoll RM. Antigen presentation by vaginal cells: role of TGFbeta as a mediator of estradiol inhibition of antigen presentation. Endocrinology 2002; 143: 2872-9.
- Kawasaki T, Choudhry MA, Suzuki T, et al. 17beta-Estradiol's salutary effects on splenic dendritic cell functions following trauma-hemorrhage are mediated via estrogen receptor-alpha. Mol Immunol 2008; 45: 376-85.
- Yang L, Hu Y, Hou Y. Effects of 17beta-estradiol on the maturation, nuclear factor kappa B p65 and functions of murine spleen CD11c-positive dendritic cells. Mol Immunol 2006; 43: 357-66.
- Bengtsson AK, Ryan EJ, Giordano D, et al. 17beta-estradiol (E2) modulates cytokine and chemokine expression in human monocyte-derived dendritic cells. Blood 2004; 104: 1404-10.
- Siracusa MC, Michael G, Overstreet MG, et al. 17 -Estradiol alters the activity of conventional and IFN-producing killer dendritic cells. J Immunol 2008; 180: 1423-31.
- Mao A, Paharkova-Vatchkova V, Hardy J, et al. Estrogen selectively promotes the differentiation of dendritic cells with characteristics of langerhans cells. J Immunol 2005; 175: 5146-51.
- Carreras E, Turner S, Paharkova-Vatchkova V, et al. Estradiol acts directly on bone marrow myeloid progenitors to differentially regulate GM-CSF or Flt3 ligand-mediated dendritic cell differentiation. J Immunol 2008; 180: 727-38.

- Kovats S, Carreras E. Regulation of dendritic cell differentiation and function by estrogen receptor ligands. Cell Immunol 2008; 252: 81-90.
- 49. Miller L, Hunt JS. Sex steroid hormones and macrophage function. Life Sci 1996; 59: 1-14.
- 50. Karpuzoglu E, Fenaux JB, Phillips RA, et al. Estrogen up-regulates inducible nitric oxide synthase, nitric oxide, and cyclooxygenase-2 in splenocytes activated with T cell stimulants: role of interferon-gamma. Endocrinology 2006; 147: 662-71.
- Soucy G, Boivin G, Labrie F, et al. Estradiol is required for a proper immune response to bacterial and viral pathogens in the female brain. J Immunol 2005; 174: 6391-8.
- 52. Suzuki S, Brown CM, Dela Cruz CD, et al. Timing of estrogen therapy after ovariectomy dictates the efficacy of its neuroprotective and antiinflammatory actions. Proc Natl Acad Sci USA 2007; 104: 6013-8.
- Naugler WE, Samurai T, Kim S, et al. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. Science 2007; 317: 121-4.

- Mor G, Sapi E, Abrahams VM, et al. Interaction of the estrogen receptors with the Fas ligand promoter in human monocytes. J Immunol 2003; 170: 114-22.
- You HJ, Kim JY, Jeong HG. 17β-Estradiol increases inducible nitric oxide synthase expression in macrophages. Biochem Biophys Res Commun 2003; 303: 1129-34.
- 56. Hayashi T, Yamada K, Esaki T, et al. Physiological concentrations of 17βestradiol inhibit the synthesis of nitric oxide synthase in macrophages via a receptor-mediated system. J Cardiovasc Pharmacol 1998; 31: 292-8.
- Ghisletti S, Meda C, Maggi A, et al. 17beta-estradiol inhibits inflammatory gene expression by controlling NF-kappaB intracellular localization. Mol Cell Biol 2005; 25: 2957-68.
- Verthelyi D. Female's heightened immune status: estrogen, T cells, and inducible nitric oxide synthase in the balance. Endocrinology 2006; 147: 659-61.