Introduction

Systemic lupus erythematosus (SLE) is an autoimmune disease which manifests with the loss of tolerance for self-antigens and the production of auto-antibodies. Autoantibodies form immune complexes which accumulate in different organs, leading to inflammation and damage. Systemic lupus erythematosus in 50-80% of cases involves the kidney, causing lupus nephritis (LN), mainly in the form of glomerulonephritis. Glomerular inflammation is followed by the activation of tubular epithelial cells, interstitial fibrosis and eventual renal failure [1].

Glomerular lesions in SLE are classified by the International Society of Nephrology/Renal Pathology Society (ISN/RPS) system or the older World Health Organization paper renal interstitial mast cell count is significantly higher in membranoproliferative glomerulonephritis than in class IV lupus nephritis

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Lupus nephritis (LN) is a severe manifestation of systemic lupus erythematosus; in LN class IV morphologic lesions may be similar to the lesions in primary membranoproliferative glomerulonephritis (MPGN). The aim of the study was to compare the counts of tryptase-positive and chymase-positive mast cells between LN class IV and MPGN.

The material consisted of 61 renal biopsies: 32 with lupus nephritis class IV, and 29 with membranoproliferative glomerulonephritis. Chymase- and tryptase-positive cells were stained by immunohistochemistry and subsequently counted. The mean count of chymase-positive mast cells was 21.94 for the whole group, 12.66 for LN class IV and 32.18 for MPGN. The mean count of tryptase-positive cells was 34.94 hpf for the entire group, 22.98 for LN class IV and 48.13 for MPGN. The differences between lupus nephritis and membranoproliferative glomerulonephritis were significant both for chymase- and tryptase-positive cells. Both chymase-positive MC counts and tryptase-positive MC counts correlated with relative interstitial volume (RIV) (R = 0.35 and R = 0.28, respectively) and with creatinine level (R = 0.35 and R = 0.43, respectively). There was also a significant correlation between age, creatinine level and RIV (R = 0.28 and R = 0.26, respectively).

Key words: SLE, lupus nephritis, membranoproliferative glomerulonephritis, mast cell, chymase, tryptase.
Organization (WHO) system; LN is divided into six classes with different mesangial, endothelial and epithelial injury patterns. Systemic lupus erythematosus class IV, also called diffuse lupus nephritis, is an endo- or extracapillary glomerulonephritis with diffuse subendothelial immune deposits, with or without mesangial alterations, which can be segmental or global [2].

Membranoproliferative glomerulonephritis (MPGN) is a rare renal disease characterized by diffuse mesangial cell proliferation, along with structural changes in glomerular capillary walls, specifically subendothelial and subepithelial deposits [3, 4]. This pattern is similar to that seen in lupus nephritis ISN/RPS class IV. Membranoproliferative glomerulonephritis can be divided into types I, II (dense deposit disease (DDD)) and III. The relationship between these three subtypes remains uncertain, but DDD is generally regarded as distinct, whereas types I and III are regarded as related entities. Types II and III are rare, while type I shows the highest morphologic similarity to proliferative lupus nephritis; thus from this point onwards, if not otherwise specified, the term MPGN refers to type I only [5].

Mast cells (MCs) are bone marrow-derived cells well known for their contribution to anaphylactic reactions, where they induce inflammation by releasing a range of mediators [6]. They are also known to participate in renal interstitial inflammation, where they contribute to renal fibrosis [7, 8]. Human mast cells contain granules with two proteases — chymase and tryptase. Mast cells can be divided into two classes, chymase-positive and chymase-negative, due to the fact that only a portion of mast cells are positive for chymase [9-11]. Chymase has the ability to convert angiotensin I to angiotensin II, which regulates intrarenal blood flow, and also participates in renal interstitial fibrosis [11, 12]. It is involved in renal remodeling by increasing the expression of transforming growth factor β (TGF-β) [13]. Tryptase is a serine protease expressed almost exclusively in mast cells. It has many biological functions, such as its role as a mitogen for fibroblasts, smooth muscle cells and bronchial epithelial cells. Additionally, it has been shown that tryptase participates in skin, lung and renal fibrosis by promoting the proliferation of fibroblasts and the production of extracellular matrix [14]. It is also known for its contribution to acute renal transplant rejection and chronic allograft nephropathy [15].

The aim of the study was to compare the counts of chymase-positive and tryptase-positive mast cells in LN ISN/RPS class IV and MPGN, two different diseases with similar morphologic changes. We examined the relationship between mast cell interstitial infiltration, renal function and diagnosis.

**Material and methods**

The material consisted of 61 renal biopsies from 32 patients with LN and 29 patients with MPGN, collected from the archives of the Department of Pathomorphology (Collegium Medicum, Jagiellonian University, Krakow). Only representative biopsies were used for this study.

The material was fixed in formalin, processed by the routine method and embedded in paraffin. 4 μm sections were cut from paraffin blocks. Sections were stained with standard hematoxylin and eosin, periodic acid-Schiff, Masson’s trichrome, Congo red, and Jones’ silver methods for histologic classification.

Primary anti-tryptase antibody (1 : 100; Leica Biosystems GmbH, Wetzlar, Germany) and anti-chymase antibody (1 : 100; Abcam, Cambridge, UK) were used. The immunohistochemistry was performed according to the routine manual method. The Lab Vision detection system (Thermo Fisher Scientific, Waltham, MA, USA) was used with 3-amino-9-ethylcarbazole as the chromogen. The slides were coun-
Interstial mast cells in different types of nephritis

interstained with Mayer hematoxylin (Thermo Fisher Scientific) and coverslipped. The immunostained slides were examined on an Olympus CH20 optical microscope equipped with a 40× lens; the number of immunopositive cells and the number of fields of view were documented. The results were expressed as the sum of positive cells from 10 high power fields, while the area of one high power field was 0.159 mm². The person performing the assessment was aware of neither the clinical data nor the diagnoses. The values of serum creatinine and daily proteinuria were obtained from the patients’ records.

For the assessment of relative interstitial volume (RIV), Masson’s trichrome stained sections were used with a Zeiss Standard 25 (Zeiss GmbH, Oberkochen, Germany) microscope equipped with a 121-point rectangular eyepiece grid (Pyser-SGI Ltd., Edenbridge, UK) and a 40× lens. The well-preserved area of the renal cortex was scanned while noting both the number of fields of view and the number of grid cross sections hitting the renal interstitium. Vessels larger than arterioles and glomeruli were ignored. The results were expressed as the ratio of the points sampled in the interstitium to the total number of points sampled in the renal cortex.

The statistical analysis was performed with Statistica 10 (StatSoft Inc., Tulsa, OK, USA). The Mann-Whitney U test was used, while correlations were measured by Spearman’s method. The level of significance was set at 0.05.

Results

The mean age of all the patients was 37.52 ± 13.83 years (all values expressed as the arithmetic mean ± SD), with a range of 19-69 years. For LN class IV, the mean age was 32.65 ± 10.53, while in MPGN it was 42.9 ± 15.18 years. The mean age for females was 35.17 ± 12.41 years and 42.35 ± 15.61 years for males. These differences were statistically significant.

The mean creatinine serum level was 99.45 ± 52.18 μmol/l, with a range of 34-310 μmol/l. In lupus nephritis class IV, the mean creatinine level was 87.3 ± 38.53 μmol/l, and in MPGN it was 113.84 ± 62.51 μmol/l. The mean daily proteinuria level was 4.77 ± 3.19 g/d, with a range of 0.11 to 14.5 g/d. In lupus nephritis class IV, the mean daily proteinuria was 4.67 ± 3.45 g/d and in MPGN it was 4.89 ± 2.93 g/d. The differences between the groups were statistically non-significant.

In both groups the mean relative interstitial volume (RIV) was 0.2 ± 0.09, with a range of 0.03 to 0.47. In class IV of lupus nephritis, the mean RIV was 0.19 ± 0.08 and in MPGN the mean RIV was –0.22 ± 0.1. The differences between the groups were statistically non-significant.

The mean count of chymase-positive MCs was 21.94 ± 22.88 per 10 high power fields, while the mean count for tryptase-positive MCs was 34.94 ± 29.29. The average ratio of chymase-positive to tryptase-positive cells was 0.69 ± 0.39.

The differences between groups in chymase-positive and tryptase-positive cell counts were statistically significant (p < 0.000047 and p < 0.000028, respectively). The differences in the ratio of chymase-positive to tryptase-positive cells between groups were not significant; however, there was a significant negative correlation between the ratio of chymase-positive to tryptase-positive cells and the chymase-positive cell count (R = –0.36). Chymase-positive MC counts and tryptase-positive MC counts were strongly correlated (R = 0.8). Both chymase-positive MC counts and tryptase-positive MC

![Fig. 3. Number of chymase positive cells in lupus nephritis class IV and membranoproliferative glomerulonephritis](image)

![Fig. 4. Number of tryptase positive cells in lupus nephritis class IV and membranoproliferative glomerulonephritis](image)
IgA nephropathy, amyloid, renovascular ischemia, in the kidney increases in renal diseases, namely di-of B-cells, T -cells, macrophages, dendritic cells and of SLE is the infiltration of immune cells, consisting tissue remodeling [16]. Also key to the pathogenesis mediators, leading to interstitial inflammation and activate other renal cells to release proinflammatory ent. In SLE class IV, deposits are located mostly in activation of the complement pathway is also pres-
nephritis is basically an immune complex disease; and fibrosis, culminating in renal failure [10]. Lupus merular lesions lead to renal interstitial inflammation other types of primary glomerulonephritis, the glo-
2BbP , activating the terminal pathway [3, 4]. Like in lizes the properdin-dependent C5 convertase (Cb3) by slow-acting nephritic factor (NeFt), which stabi-
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lizes the properdin-dependent C5 convertase (Cb3) 2BbP, activating the terminal pathway [3, 4]. Like in other types of primary glomerulonephritis, the glo-
merular lesions lead to renal interstitial inflammation and fibrosis, culminating in renal failure [10]. Lupus nephritis is basically an immune complex disease; activation of the complement pathway is also present. In SLE class IV, deposits are located mostly in the subendothelial and mesangial areas, where they activate other renal cells to release proinflammatory mediators, leading to interstitial inflammation and tissue remodeling [16]. Also key to the pathogenesis of SLE is the infiltration of immune cells, consisting of B-cells, T-cells, macrophages, dendritic cells and mast cells [17-22]. The number of mast cells found in the kidney increases in renal diseases, namely di-
abetic nephropathy, acute renal allograft rejection, IgA nephropathy, amyloid, renovascular ischemia, reflux nephropathy, polycystic kidney disease and drug-induced nephropathy [7, 10]. Little information is available with regards to the role of mast cells in LN and membranoproliferative glomerulonephritis and to the contribution of these cells to the pathogenesis of these diseases. Togawa et al. investigated chymase expression in renal biopsies from children with inter alia MPGN. The study showed that the number of chymase-pos-
itive mast cells correlates with the mean intensity of the interstitial fibrosis, which is similar to the results obtained in our study [12]. Danilewicz et al. analyzed the relationship between mast cell count and the expression of interstitial al-
pha-smooth muscle actin (SMA), along with the inter-
stitial inflammatory infiltrates in MPGN type I. They found significant positive correlations between inter-
stitial tryptase-positive cells and interstitial expression of SMA, interstitial volume, serum creatinine as well as CD 43+ and CD 68+ cells, which suggests the role of MCs in the development of interstitial fibrosis [23]. The same group conducted an analogous study with rebiopsied patients with MPGN, where they ob-
tained similar results [24]. Danilewicz et al. demon-
strated in another study that the number of interstitial tryptase-positive cells was significantly higher in class V lupus nephritis than in idiopathic membranous glo-
merulopathy. Both groups exhibited significant posi-
tive correlations between mast cell count and relative interstitial volume (RIV), along with serum creatinine level [25]. In our study, more mast cells were seen in MPGN than in LN. In our present and previous studies [26], we have found the same positive correlation with RIV and creatinine level. Hiromura et al. ex-
amed the number of interstitial mast cells in both MPGN and LN. They found a correlation between mast cell count, interstitial fibrosis, creatinine level and leukocyte infiltration, but not with urinary pro-	ein excretion; the same results were obtained in our study [26]. In our previous study, we found that mast cell counts differ across classes of SLE, which suggests that these cells play a role in the pathogenesis of LN [26]. We have shown that despite the similar pattern of pathological changes in the renal glomerulus in LN class IV and MPGN, the quantity of mast cells in the renal interstitium is different. We also confirmed that mast cells contribute to the tubulointerstitial fibrosis, while their numbers correlate with RIV. This necessi-
tates further investigation to explain the role of mast cells in the pathogenesis of and MPGN.

Discussion

Systemic lupus erythematosus class IV and MPGN are two entities differing in etiology and biology, but in some cases they share a similar morphologic pattern. In our study we examined the relationship between the type of renal disease and number of MCs found in the renal tissue. Interestingly, more mast cells were seen in MPGN than in LN class IV, both for chymase-positive and tryptase-positive mast cells. Additionally, the counts of mast cells correlated with relative interstitial volume (RIV), which confirms the role of mast cells in renal interstitial fibrosis.

Membranoproliferative glomerulonephritis displays a pattern of glomerular injury seen in both idiopathic forms and secondary diseases associated with systemic or infectious disorders such as hepatitis C or LN. The pathogenesis of MPGN is complex and not clearly understood. The pathogenesis is thought to be different in each subtype of MPGN, but the comple-
ment system is significantly involved in all types regardless. Type I is probably due to the accumulation of circulating immune complexes which trigger the complement cascade via the classical pathway. In MPGN type II, the complement system is activat-
ed by the alternative pathway, due to the activity of C3 nephritic factor (C3NeF); this IgG autoantibody prevents the inactivation of C5 convertase (C3bBb) of the alternative pathway, resulting in the break-
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merular lesions lead to renal interstitial inflammation and fibrosis, culminating in renal failure [10]. Lupus nephritis is basically an immune complex disease; activation of the complement pathway is also present. In SLE class IV, deposits are located mostly in the subendothelial and mesangial areas, where they activate other renal cells to release proinflammatory mediators, leading to interstitial inflammation and tissue remodeling [16]. Also key to the pathogenesis of SLE is the infiltration of immune cells, consisting of B-cells, T-cells, macrophages, dendritic cells and mast cells [17-22]. The number of mast cells found in the kidney increases in renal diseases, namely di-
abetic nephropathy, acute renal allograft rejection, IgA nephropathy, amyloid, renovascular ischemia,

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